MERKEL CELL CARCINOMA: ADVANCES IN PATHOGENESIS, DIAGNOSIS AND PROGNOSTIC FACTORS

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Rezumat

Carcinomul cu celule Merkel (CCM) este o tumoră neuroendocrină a pielii cu grad ridicat de agresivitate, caracterizându-se prin o probabilitate mare de recurență și metastazare. Cu toate că aceasta este considerată o formă rară de cancer, recent s-a observat o creștere rapidă a incidenței. De aceea lumea de specialitate depune eforturi sporite în definirea ei. Articolul de față tratează atât aspectele cunoscute ale acestei tumori, cât și aspecte controversate legate de patogenia, diagnosticul și factorii de prognostic ai CCM.

Cuvinte cheie: Carcinom cu celule Merkel, Merkel poliomavirus celule, căi oncogenice, imunohisto-chimie, prognostic.

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Summary

Merkel cell carcinoma (MCC) is a highly aggressive neuroendocrine skin tumor, characterized by high incidence of recurrences and metastases. It is a relatively unusual form of cancer however, its incidence is rapidly increasing. Therefore, the interest in defining this disease has escalated rapidly. This article addresses what is known and what is still controversial about the pathogenesis, diagnosis and prognostic factors in Merkel cell carcinoma.

Keywords: Merkel cell carcinoma, Merkel cell polyomavirus, oncogenic pathways, immunohistochemistry, prognosis.

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Introduction

Merkel cell carcinoma (MCC) is a highly aggressive neuroendocrine skin cancer, characterized by high incidence of local recurrences, regional nodal and distant metastases. Mortality rates are estimated to 33% within 5 years, the highest of any dermatological cancer [1]. It is a relatively uncommon form of cancer however its incidence had increased in the last years which had triggered a higher interest in better defining the pathogenesis of this disease and searching for new therapies. MCC has been first described in

1972 by Toker and named trabecular carcinoma [2]. Later studies found neurosecretory granules in the cytoplasm of tumor cells, similar to those seen in non-neoplastic Merkel cells (MC) which resulted in renaming the tumor as MCC.

Although it is believed that MCC originates from MC, there is some controversy regarding the "cell-of-origin". The traditional view is sustained by a similar (but not identical) immunophenotype. It is also generally accepted that MCs are of neuroendocrine origin, being the only cutaneous cells that form electron-dense neurosecretory granules, a histologic feature also

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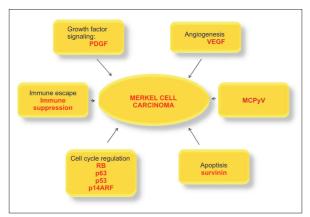


Fig. 1. Main factors implicated in the pathogenesis of Merkel cell carcinoma

discovered in MCC. Some authors advance the hypothesis that MCC originates in the pluripotent epidermal stem cells. This idea is supported by the squamous differentiation observed in certain cases of MCC and its metastases and by the frequent association of MCC with squamous cell carcinoma [6]. Recently, both ideas have been linked by the proof that mammalian MCs develop from epidermal stem cells, rather than from neural crest progenitors [3-5].

The pathogenesis of MCC is not completely elucidated. Frequent associations with exposure to ultraviolet (UV), ionizing, and infrared radiation have been reported [6-8]. Many studies in the literature have described the association between MCC and iatrogenic immunosuppresion, the tumor being noted in almost every autoimmune disease, with an increasing incidence with the use of tumor necrosis factor (TNF) inhibitors [9]. MCC has also been reported after organ transplantation, especially in renaltransplant recipients. These observations support the idea that chronic immunosuppression increases the risk of MCC [10-11]. One of the most important step in understanding the etiology of the disease was the discovery of a novel polyomavirus in MCC tumor specimens [12]. Several oncogenic pathways have also been studied [13-16]

The clinical presentation of this tumor is nonspecific. It is usually described as a painless indurated pink to reddish-blue or brown intradermal nodule with a rapid growth, on the sun-exposed areas of elderly. The unspecific manifestations, mimicking many benign and

malignant skin tumors make MCC difficult to diagnose clinically before biopsy. The most common presumptive diagnosis at early stages is cyst. Thus, the diagnosis is made by histopathologic examination and immunohistochemistry [3,17].

Various factors have been described as indicators of a poor prognosis. In the current paper, we review the advances in pathogenesis diagnosis, and potential prognostic factors. (Fig. 1)

Merkel cell polyomavirus and its role in pathogenesis

As mentioned before, MCC was clearly associated with UV exposure and immuno-suppression but little was known about the pathological mechanisms involved in tumorigenesis. However, Feng et al [12] revolutionized our understanding of this tumor by discovering a novel viral agent that integrates in the genome of MCC tumor cells: Merkel cell polyoma virus (MCPyV). Further studies demonstrated that viral integration preced clonal expansion, MCPyV being found at distinct sites in different MCCs [18].

MCPyV is a member of Polyomaviridae family, now constituted as a single genus (Polyomavirus). It is a 40 nm non-enveloped double stranded DNA viruse, whose genome consists of approximately 4700-5400 base pairs and codes for 3 structural proteins and few early and late proteins. Early proteins, particularly small and large T antigens, promote genome replication and tumorigenesis, while the late region encodes viral capsid proteins [19]. Large T antigen has the ability to bind DNA polymerase, primase, topoisomerase I and tumor suppressor proteins, such as retinoblastoma protein (pRb) and p53. This antigen seems to possess features commonly seen in oncogenic polyomaviruses, such as the LxCxE motif which can directly bind pRb. Large T antigen also possesses a helicase motif required for virus replication. MCPyV small T cell antigen has the property to inactivate protein phosphatase 2A. Upon integration into the host genome, MCPyV DNA generally harbors tumor-specific mutations, including deletion or truncation at carboxyl-terminal half of the large T antigen... Taken together, these findings lend support to the hypothesis that MCPyV viral proteins are involved in oncogenesis [20-22].

However, approximately 20% of MCCs are negative for this virus, so it is not "necessary" for tumorigenesis. From another point of view, only a small proportion of people infected with MCPyV develop the disease [23]. It was suggested that MCPyV-negative MCCs have fewer genomic deletions than MCPyV-positive MCCs [24]. Added to this, it was highlighted that expression of p53 and KIT was accompanied by the absence or low copy numbers of MCPyV DNA [25].

Seropositivity rates for antibodies against viral capsid proteins 1 (VP1) range from 40 to 88% [26-29]. Despite this relatively high prevalence, the antibody titers are higher in patients with MCC than in general population and seem to correlate with the presence of MCPyV in the skin [30-31]. In contrast, there was no correlation between tumor characteristics and viral load or antibody titer. However, higher antibody titers seem to improve the prognosis, suggesting a strong immune response [32]. A number of studies have shown that MCPvVpositive cases tend to have a better clinical outcome than the negative ones [33,34], while other studies did not find any differences [35]. Cases with MCPyV-positive tumors and an aggressive course have also been described [36].

Oncogenic pathways

The most important processes involved in carcinogenesis are the activation of growth-stimulating oncogenes and inactivation of tumor suppressor genes. However, many studies revealed little involvement of several known oncogenic pathways such as: antiapoptotic Bcl-2 family, Wnt pathway, MAP kinase signaling pathway [37]. PTEN does not seemingly have an important role in MCC oncogenesis, although loss of heterozygosity is a frequent event [38]. Mutations in other tumor suppressor genes such as p73 and CDKN2A were rarely observed in MCC [39,40]. Wnt pathway has also been evaluated, but mutations in β-catenin, APC, AXIN1 or AXIN2 were not identified [41].

Although several studies reported that p53 is not a key protein in the pathogenesis of MCC, Lassacher et al [14] suggested the involvement of p14ARF/mdm2/p53 pathway in a subset of MCCs. The authors demonstrated hyperme-

thylation in the promoter region of p14ARF in 42% of cases, resulting in an altered expression pattern of the gene. The classical mitogenactivated protein (MAP) kinase signaling pathway, one of the most studied pathways in oncogenesis, has a central role in proliferation, cell-cycle arrest, suppresion of apoptosis, migration and teminal differentiation. It is activated in several cancers through mutations involving of tyrosine kinase receptors (EGFR, KIT, HER2), RAS family (H-RAS, K-RAS, N-RAS), and B-RAF [42]. However, several studies showed negativity for H-Ras, K-Ras, N-Ras [13]. B-RAF, [43], EGFR, and Her2 expression [44].

In contrast, KIT protein, a receptor belonging to the PDGFR family, is overexpressed in 59-95% of MCCs. KIT protooncogene, located on chromosome 4p, encodes 976 amino-acid protein (CD117 or KIT) containing an extracellular, transmembrane, juxtamembrane and tyrosine kinase domain. KIT activation is followed by processes like receptor dimerization and internalization, substrate phosphorylation and autophosphorylation, activation of protein kinases and phospholipases and transcription of different proto-oncogenes. Mutations in gene have been identified in several tumors such as gastrointestinal stromal tumors (GISTs), mast cell neoplasms or melanoma and are believed to play a central pathogenetic role [45-48]. Activating mutations of KIT gene in GISTs were identified in exon 11 encoding the juxtamembrane domain, exons 13 and 17 that encode the tyrosine kinase domain, and less frequently in exon 9 enconding the extracellular domain [49,50]. Investigations of the same exons have failed to find any mutations in MCCs [48,51]. More recently, activating mutations of the platelet-derived growth factor receptor alpha have been identified in 33% of MCC cases [52]. Activation of the PI3K/AKT signaling pathway represents one of the most frequent events in human cancer. Oncogenic activity is sustained by the identification of somatic mutations in the PIK3CA gene in a wide range of tumors [53-54]. Oncogenic mutations have also been noted in certain domains of AKT1[55]. Hafner et al [56] included MCC in the list of cancers harboring PIK3CA mutations. The authors demonstrated that AKT phosphorylation and

PI3K/AKT pathway activation was not correlated with the presence of MCPyV and oncogenic PIK3CA mutations in MCC. The low frequency of PIK3CA mutations indicates that additional mechanisms contribute to the activation of this pathway in MCC. Additionally, mutations in ATOH1, a tumour suppressor gene encoding a transcription factor involved in Merkel cell differentiation, have been reported [57].

Histologic parameters and prognosis

The typical histological finding is a dermal tumor with extensions to underlying subcutaneous tissue. The epidermis, papillary dermis and adnexal structures are usually spared. Apoptotic cells and mitotic figures are frequently seen. The small blue cells with sparse cytoplasm are characteristic. The nuclei are hyperchromatic without prominent nucleoli and there is a very high nuclear cytoplasmic ratio. The chromatin is displayed in typical salt and pepper pattern. Histologic variants include a large cell type (with a trabecular pattern), an intermediate-cell type, and a small-cell type with resemblance to skin metastases of small-cell lung carcinoma (SCLC). If the architecture is considered, MCC are classified into a solid or organoid variant, a diffuse variant, and a trabecular variant (the least common pattern) [58].

Regarding tumor staging, low stage predicts a relatively good prognosis, but it is not an ideal indicator as a significant proportion of patients with stage I still die within 5 years from diagnosis [59]. Furthermore, five inconsistent staging systems have been proposed during the past 20 years [60]. Thus, alternative independent pathologic markers of prognosis are helpful, especially for low stage tumors,. Tumor size has been found to correlate with survival in clinical stage I, but it loses significance in higher stages [61,62]. Sandel et al [63] found no correlation between tumor size and prognosis.

Histologic features associated with a poor prognosis include a mitotic rate of more than 10 per high power field, small cell size, and a diffuse growth pattern. Other studies failed to find any association between prognosis and tumor pattern or mitotic rate. Regarding the depth of invasion, not all studies agree. Average tumor thickness associated with the risk of

distant metastases has been reported in MCC to be close to 10 mm, although other studies have found no correlation between tumor thickness and overall survival. The extension of the tumor into the underlying subcutaneous tissue was correlated with a shortened survival, but another study failed to confirm this finding. It was also suggested that mast cell infiltrates have negative prognostic value in MCC. Lymphovascular invasion was found to affect the prognosis in one study, but in other studies it was not significantly associated with prognosis. The relation between the associated inflammatory infiltrate and prognosis is contradictory reported. Other histological markers such as ulceration, tumor necrosis, or apoptosis did not reveal any correlation with prognosis [62-67].

Immunohistochemistry: diagnostic and prognostic tool

The diagnosis of MCC is challenging, often being confused with benign conditions such as cyst, lipoma, dermatofibroma or vascular lesions [68]. Therefore, the immunohistochemistry is necessary for diagnosis. The tumour expresses both neuroendocrine and epithelial markers. Neuroendocrine differentiation is sustained by the positivity for neuronspecific enolase (NSE), chromogranin A (CrA) and synaptophysin. These markers show a diffuse cytoplasmic pattern. The tumor can also express other neural markers such neurofilaments, microtubule-associated protein (MAP) 2, CD56, and CD57. S-100 is negative in most cases [69]. CK20 is the most important epithelial marker, the paranuclear dotlike staining being characteristic. MCC is also frequently positive for other epithelial markers, including AE1/AE3, CAM5.2, and Ber-EP4 [70]. CK7 is rarely positive, and a few CK7+/CK20cases have been reported [71]. (Table 1)

MCCs share many histological features with small cell carcinomas of other organs, including

Table 1. Positive IHC markers in MCC

Neuroendicrine markers	Epithelial markers
NSE	CK20
CrA	AE1/AE3
Synaptphysin	Ber-EP4
MAP2	CK7 (rarely)
CD56	
CD57	

SCLC [72]. For this reason, IHC plays an essential role for differentiation between these entities. CK7/CK20 staining patterns are useful in some cases. A significant proportion of MCC show a CK7(-)CK20(+) pattern, whereas some of the small cell carcinomas of the lung are CK7(+)CK20(-). There are lots of cases of both entities that express neither CK7 nor CK20. Even CK7(+)CK20(-) or CK7(+)CK20(+) MCC cases have been reported [72,73]. Therefore, thyroid transcription factor-1 (TTF-1), a tissue specific transcription factor expressed in epithelial cells of the thyroid, lung and brain, is a very useful marker for differentiation. Combining TTF-1 with CK-20 offers a strong basis for diagnosis. MCC is CK-20 positive and TTF-1 negative, while SCLC is TTF-1 positive [74-79]. Ralston et al [80] proposed MASH1 (achaete-scute complex-like 1, ASCL1) for discriminating between the two conditions. According to their study, 83% of small cell carcinomas of the lung expressed MASH1, and only 73% expressed TTF-1. MASH1 was negative in all MCC, while TTF-1 was expressed in 3% of MCC. MASH1 can be used together with TTF-1, CK7 and CK20 in a panel of immunostains, to facilitate the diagnosis [81]. A monoclonal antibody (CM2B4) can identify MCPyV-positive MCCs with diffuse nuclear staining in immunohistochemistry, approximately 80% sensitivity compared with polymerase chain reaction [82].

Immunohistochemistry can also be to investigate the prognosis in MCC. To date, various prognostic markers have been reported. However, the lack of standardized methods makes the results considerably variable, only a few markers sustaining their prognostic value [70].Ki-67, the most useful marker of cell proliferation, correlates with evolution towards metastasis and poor prognosis in various studies [65,82,83]. Although p53 was reported to correlate with a poor prognosis [25,84], this finding was not confirmed by others [83,85,86]. Another IHC marker with prognosis value is the p63 protein, a member of the p53 family of transcription factors. Asioli et al [86], demonstrated an aggressive course, with poor overall survival in patients with p63 positive tumors. Survivin, an antiapoptosis protein, is expressed in most cancers, including MCC, while normal cells lack its expression. It was shown that nuclear expression was correlated with an aggressive course of MCC, whereas a predominantly cytoplasmic staining correlated with disease-free survival [87]. A differential subcellular division of survivin was correlated with prognosis. The authors suggest that survivin staining can provide important information concerning the possibility for therapeutic intervention, contrasting with other prognostic markers [88].

As mentioned above, a significant percentage of MCCs express KIT, but it was not correlated with prognosis in most studies. [45,47,65,85] Nevertheless, Andea et al. demonstrated that a high KIT expression may contribute to poorer survival rates [89]. KIT expression was also associated with lymphovascular invasion and high mitotic rate, suggesting a more aggressive course [47].

Overexpression of a large number of other IHC markers have been associated with metastasis: vascular endothelial growth factor (VEGF), p38, matrix metalloproteinase (MMP) 7, MMP 10/2, tissue inhibitor of metalloproteinase 3, stromal NF-kappa B, synaptophysin, CD44, and CD151 [.90-92]. In contrast, CD9 expression was significantly correlated with better overall survival [92] (Table 2)

Table 2. IHC markers for poor prognosis and metastasis

Ki-67	
p63	
Nuclear expresion of survivin	
high KIT expresion	
VEGF	
p38	
MMP 7 and MMP 10/2	
tissue inhibitor of metalloproteinase 3	
stromal NF-kappa B	
Synaptophysin	
CD44	
CD151	

Conclusions

Increasing incidence and mortality of MCC make the early diagnosis and identification of prognostic factors mandatory. Viral and cytogenetic studies represent an exciting direction for future studies to determine the oncogenic events involved in promotion or maintenance of MCC. Furthermore, the understanding of characteristic signal transduction pathways will provide opportunities to explore new targeted therapies for this aggressive tumor.

Bibliografie/Bibliography

- 1. Hodgson NC.Merkel cell carcinoma: changing incidence trends. J Surg Oncol 2005;89:1-4.
- 2. Toker C. Trabecular carcinoma of the skin. Arch Dermatol 1972;105:107–10.
- 3. Dinh V, Feun L, Elgart G, Savaraj N. Merkel cell carcinomas. Hematol Oncol Clin North Am 2007;21:527-44.
- 4. Werling AM, Doerflinger Y, Brandner JM, et al. Homo- and heterotypic cell-cell contacts in Merkel cells and Merkel cell carcinomas: heterogeneity and indications for
- 5. Tilling T, Moll I. Which are the cells of origin in merkel cell carcinoma? J Skin Cancer. 2012;2012:680410.
- 6. Lunder EJ, Stern RS. Merkel-cell carcinomas in patients treated with methoxsalen and ultraviolet A radiation. N Engl J Med.1998;339:1247-8.
- 7. Hewitt JB, Sherif A, Kerr KM, et al. Merkel cell and squamous cell carcinomas arising in erythema ab igne. *Br J Dermatol* 1993;128:591-92.
- 8. Jones CS, Tyring SK, Lee PC, et al. Development of neuroendocrine (Merkel cell) carcinoma mixed with squamous cell carcinoma in erythem ab igne. *Arch Dermatol* 1988;124:110-3.
- 9. Stone JH, Holbrook JT, Marriott MA, et al. Solid malignancies among patients in the Wegener's Granulomatosis Etanercept Trial. Arthritis Rheum 2006;54: 1608-18.
- 10. Urbatsch A, Sams WM, Urist MM, et al. Merkel cell carcinoma occurring in renal transplant patients. *J Am Acad Dermatol* 1999;41:289-91.
- 11. Penn I, First MR. Merkel's cell carcinoma in organ recipients: report of 41 cases. Transplantation 1999;68:1717-21.
- 12. Feng H. Shuda M, Chang Y, et al. Clonal Integration of a Polyomavirus in Human Merkel Cell Carcinoma. *Science* 2008;319:1096–100.
- 13. Popp S, Waltering S, Herbst C, et al. UV-B-type mutations and chromosomal imbalances indicate common pathways for the development of Merkel and skin squamous cell carcinomas. Int J Cancer 2002;99:352-60.
- 14. Lassacher A, Heitzer E, Kerl H, et al. p14ARF hypermethylation is common but INK4a-ARF locus or p53 mutations are rare in Merkel cell carcinoma. J Invest Dermatol 2008;128:1788-96.
- 15. Becker JC, Schrama D, Houben R. Merkel cell carcinoma. Cell Mol Life Sci 2009;66:1-8. Houben R, Schrama D, Becker JC. Molecular pathogenesis of Merkel cell carcinoma. Exp Dermatol 2009;18:193-8.
- 16. Helmbold P, Lahtz C, Enk A, et al. Frequent occurrence of RASSF1A promoter hypermethylation and Merkel cell polyomavirus in Merkel cell carcinoma. Mol Carcinog 2009;48:903-9.
- 17. O'Connor WJ, Brodland DG. Merkel cell carcinoma. Dermatol Surg. 1996;22:262-7.
- 18. Sastre-Garau X, Peter M, Avril MF, et al. Merkel cell carcinoma of the skin: pathological and molecular evidence for a causative role of MCV in oncogenesis. *J Pathol* 2009;218:48-56.
- 19. Feng H, Kwun HJ, Liu X, et al. Cellular and viral factors regulating Merkel cell polyomavirus replication. *PLoS One* 2011;6:e22468.
- 20. Garneski KM, DeCaprio JA, Nghiem P. Does a new polyomavirus contribute to Merkel cell carcinoma? *Genome Biol* 2008;9:228.
- 21. Houben R, Shuda M, Weinkam R, et al. Merkel cell polyomavirus-infected Merkel cell carcinoma cells require expression of viral T antigens. *J Virol* 2010;84:7064-72.
- 22. Shuda M, Feng H, Kwun HJ, et al. T antigen mutations are a human tumor-specific signature for Merkel cell polyomavirus. Proc Natl Acad Sci U S A 2008;105:16272-7.
- 23. Schowalter RM, Pastrana DV, Pumphrey KA, et al.)Merkel Cell Polyomavirus and Two Previously Unknown Polyomaviruses Are Chronically Shed from Human Skin. *Cell Host & Microbe* 2010;6:509–15.
- 24. Paulson KG, Lemos BD, Feng B, et al. Array-CGH reveals recurrent genomic changes in Merkel cell carcinoma including amplification of L-Myc. *J Invest Dermatol* 2009;129:1547–55.
- 25. Waltari M, Siĥto H, Kukko H, et al. Association of Merkel cell polyomavirus infection with tumor p53, KIT, stem cell factor, PDGFR-alpha and survival in Merkel cell carcinoma. *Int J Cancer* 2011;129:619–68.
- 26. Kean JM, Rao S, Wang M, et al. Seroepidemiology of human polyomaviruses. PLoS Pathog 2009;5:e1000363.
- 27. Carter JJ, Paulson KG, Wipf GC, et al. Association of Merkel cell polyomavirus-specific antibodies with Merkel cell carcinoma. *J Natl Cancer Inst* 2009;101:1510–22.
- 28. Pastrana DV, Tolstov YL, Becker JC, et al. Quantitation of human seroresponsiveness to Merkel cell polyomavirus. *PLoS Pathog* 2009; 5:e1000578.
- 29. Viscidi RP, Rollison DE, Sondak VK, et al. Age specific-seroprevalence to Merkel cell polyomavirus, BK virus and JC virus. Clin Vaccine Immunol 2011;18:1737–43.
- 30. Faust H, Pastrana DV, Buck CB, et al. Antibodies to Merkel cell polyomavirus correlate to presence of viral DNA in the skin. *J Infect Dis* 2011; 203:1096–1100.
- 31. Pastrana DV, Wieland U, Silling S, et al. Positive correlation between Merkel cell polyomavirus viral load and capsid-specific antibody titer. *Med Microbiol Immunol* 2012;201:17-23.
- 32. Touze A, Le BE, Laude H, et al. High levels of antibodies against merkel cell polyomavirus identify a subset of patients with merkel cell carcinoma with better clinical outcome. *J Clin Oncol* 2011; 29:1612–9.

- 33. Sihto H, Kukko H, Koljonen V, et al. Clinical factors associated with Merkel cell polyomavirus infection in Merkel cell carcinoma. *J Natl Cancer Inst* 2009;101:938-45.
- 34. Bhatia K, Goedert JJ, Modali R, et al. Immunological detection of viral large T antigen identifies a subset of Merkel cell carcinoma tumors with higher viral abundance and better clinical outcome. *Int J Cancer* 2010;127:1493-6.
- 35. Schrama D, Peitsch WK, Zapatka M, et al. Merkel cell polyomavirus status is not associated with clinical course of Merkel cell carcinoma. *J Invest Dermatol* 2011;131:1631-8.
- 36. Andea AA, Patel R, Ponnazhagan S, Kumar S, DeVilliers P, Jhala D, et al. Detection of Merkel cell polyomavirus in formalin-fixed, paraffin-embedded tissue of Merkel cell carcinoma and its correlation with prognosis. *J Clin Oncol* 2009;27:6027.
- 37. Becker JC, Schrama D, Houben R. Merkel cell carcinoma. Cell Mol Life Sci 2009;66:1-8. Houben R, Schrama D, Becker JC. Molecular pathogenesis of Merkel cell carcinoma. *Exp Dermatol* 2009;18:193-8.
- 38. Van Gele M, Leonard JH, Van Roy N, et al. Frequent allelic loss at 10q23 but low incidence of PTEN mutations in Merkel cell carcinoma. *Int J Cancer* 2001;92:409–13.
- 39. Van Gele M, Kaghad M, Leonard JH, et al. Mutation analysis of *P73* and *TP53* in Merkel cell carcinoma. *Br J Cancer* 2000;82:823–6.
- 40. Koljonen V, Tukiainen E, Haglund C, et al. Cell cycle control by p21, p27 and p53 in Merkel cell carcinoma. *Anticancer Res*2006;26:2209–12.
- 41. Liu S, Daa T, Kashima K, Kondoh Y, et al. The Wnt-signaling pathway is not implicated in tumorigenesis of Merkel cell carcinoma. *J Cutan Pathol* 2007:34:22–6.
- 42. Hanahan D, Weinberg RA. The hallmarks of cancer. Cell 2000:100:57-70.
- 43. Houben R, Michel B, Vetter-Kauczok CS, et al. Absence of classical MAP kinase pathway signalling in Merkel cell carcinoma. *J Invest Dermatol* 2006:126:1135–42.
- 44. Brunner M, Thurnher D, Pammer J, et al. Expression of VEGFA/C, VEGF-R2, PDGF-alpha/beta, c-kit, EGFR, Her-2/Neu, Mcl-1 and Bmi-1 in Merkel cell carcinoma. *Mod Pathol* 2008:21:876-84.
- 45. Su LD, Fullen DR, Lowe L, et al. CD117 (KIT receptor) expression in Merkel cell carcinoma. *Am J Dermatopathol* 2002;24:289-93.
- 46. Strong S, Shalders K, Carr R, et al. KIT receptor (CD117) expression in Merkel cell carcinoma. *Br J Dermatol* 2004;150:384-5.
- 47. Feinmesser M, Halpern M, Kaganovsky E, et al. C-kit expression in primary and metastatic Merkel cell carcinoma. *Am J Dermatopathol* 2004;26:458-62.
- 48. Swick BL, Ravdel L, Fitzpatrick JE, et al. Merkel cell carcinoma: evaluation of KIT (CD117) expression and failure to demonstrate activating mutations in the C-KIT proto-oncogene implications for treatment with imatinib mesylate. *J Cutan Pathol* 2007;34:324-9.
- 49. Lasota J, Corless CL, Heinrich MC, et al. Clinicopathologic profile of gastrointestinal stromal tumors (GISTs) with primary KIT exon 13 or exon 17 mutations: a multicenter study on 54 cases. *Mod Pathol* 2008;21:476-84.
- 50. Prakash S, Sarran L, Socci N, et al. Gastrointestinal stromal tumors in children and young adults: a clinicopathologic, molecular, and genomic study of 15 cases and review of the literature. *J Pediatr Hematol Oncol* 2005:27:179-87.
- 51. Kartha RV, Sundram UN. Silent mutations in KIT and PDGFRA and coexpression of receptors with SCF and PDGFA in Merkel cell carcinoma: implications for tyrosine kinase-based tumorigenesis. *Mod Pathol* 2008;21:96-104.
- 52. Swick BL, Ravdel L, Fitzpatrick JE, et al. Platelet-derived growth factor receptor alpha mutational status and immunohistochemical expression in Merkel cell carcinoma: implications for treatment with imatinib mesylate. *J Cutan Pathol* 2008;35:197-202.
- 53. Karakas B, Bachman KE, Park BH. Mutation of the PIK3CA oncogene in human cancers. Br J Cancer 2006:94:455-9.
- 54. Hafner C, Lopez-Knowles E, Luis NM, et al. Oncogenic PIK3CA mutations occur in epidermal nevi and seborrheic keratoses with a characteristic mutation pattern. *Proc Natl Acad Sci U S A* 2007:104:13450–4.
- 55. Bleeker FE, Felicioni L, Buttitta F, et al. AKT1(E17K) in human solid tumours. Oncogene 2008:27:5648–50.
- 56. Hafner C, Houben R, Baeurle A, et al. Activation of the PI3K/AKT pathway in Merkel cell carcinoma. *PloS One* 2012;7:e31255.
- 57. Bossuyt W, Kazanjian A, De Geest N, et al. Atonal homolog 1 is a tumor suppressor gene. PLoS Biol 2009;7:e39.
- 58. Koljonen VS. Merkel cell carcinoma. World J Surg Oncol 2006;4:7. Mott RT, Smoller BR, Morgan MB. Merkel cell carcinoma: a clinicopathologic study with prognostic implications. *J Cutan Pathol* 2004;31:217-23.
- 59. Allen PJ, Bowne WB, Jaques DP, et al. Merkel cell carcinoma: prognosis and treatment of patients from a single institution. *J Clin Oncol* 2005;23:2300-9.
- 60. Lemos B, Storer B, Iyer J, et al. Pathologic nodal evaluation improves prognostic accuracy in Merkel cell carcinoma: Analysis of 5,823 cases as the basis of the first consensus staging system for this cancer. *J Am Acad Dermatol* 2010;63:751–61.

- 61. Yiengpruksawan A, Coit DG, Thaler HT, Urmacher C, Knapper WK. Merkel cell carcinoma. Prognosis and management. *Arch Surg* 1991;126:1514-9.
- 62. Mott RT, Smoller BR, Morgan MB. Merkel cell carcinoma: a clinicopathologic study with prognostic implications. *J Cutan Pathol* 2004;31:217-23.
- 63. Sandel HD, Day T, Richardson MS, et al. Merkel cell carcinoma: does tumor size or depth of invasion correlate with recurrence, metastasis, or patient survival? *Laryngoscope* 2006;116:791-5.
- 64. Skelton HG, Smith KJ, Hitchcock CL, et al. Merkel cell carcinoma: analysis of clinical histologic, and immunohistologic features of 132 cases with relation to survival. *J Am Acad Dermatol* 1997;37:734.
- 65. Llombart B, Monteagudo C, Lopez-Guerrero JA, et al. Clinicopathological and immunohistochemical analysis of 20 cases of Merkel cell carcinoma in search of prognostic markers. *Histopathology* 2005;46:622-34.
- 66. Pilotti S, Rilke F, Bartoli C, Grisotti A. Clinicopathologic correlations of cutaneous neuroendocrine Merkel cell carcinoma. J Clin Oncol 1988;6:1863-73.
- 67. Goldberg SR, Neifeld JP, Frable WJ. Prognostic value of tumor thickness in patients with Merkel cell carcinoma. *J Surg Oncol* 2007;95:618-22.
- 68. Heath M, Jaimes N, Lemos B et al. Clinical characteristics of Merkel cell carcinoma at diagnosis in 195 patients: the AEIOU features. *J Am Acad Dermatol* 2008:58:375–81.
- 69. Kuwamoto S. Recent advances in the biology of Merkel cell carcinoma. Hum Pathol 2011;42:1063-77.
- 70. Mount SL, Taatjes DJ. Neuroendocrine carcinoma of the skin (Merkel cell carcinoma). An immunoelectron-microscopic case study. *Am J Dermatopathol* 1994;16:60-5.
- 71. Pilloni L, Manieli C, Senes G, et al. Merkel cell carcinoma with an unusual immunohistochemical profile. *Eur J Histochem* 2009;53:275-8.
- 72. Shahab N, Mirza IA, Doll D. Extrapulmonary small cell carcinoma. Semin Oncol 2007;34:1-2.
- 73. Fernández-Figueras MT, Puig L, Musulen E, et al. Prognostic significance of p27Kip1, p45Skp2 and Ki67 expression profiles in Merkel cell carcinoma, extracutaneous small cell carcinoma, and cutaneous squamous cell carcinoma. Histopathology 2005;46:614-21.
- 74. Koljonen V, Tukiainen E, Haglund C, Böhling T. Proliferative activity detected by Ki67 correlates with poor outcome in Merkel cell carcinoma. Histopathology 2006;49:551-3.
- 75. Wang NP, Zee S, Bacchi CE, et al. Coordinate expression of cytokeratins 7 and 20 defines unique subsets of carcinomas. Appl Immunohistochem 1995;3:99–107.
- 76. Lazzaro D, Price M, de Felice M, et al. The transcription factor TTF-1 is expressed at the onset of thyroid and lung morphogenesis and in restricted regions of the foetal brain. *Development* 1991;113:1093-104.
- 77. Byrd-Gloster AL, Khoor A, Glass LF, et al. Differential expression of thyroid transcription factor 1 in small cell lung carcinoma and Merkel cell tumor. *Hum Pathol* 2000;31:58-62.
- 78. Cheuk W, Kwan MY, Suster S, et al. Immunostaining for thyroid transcription factor 1 and cytokeratin 20 aids the distinction of small cell carcinoma from Merkel cell carcinoma, but not pulmonary from extrapulmonary small cell carcinomas. *Arch Pathol Lab Med* 2001;125:228-31.
- 79. Ordonez NG. Value of thyroid transcription factor-1 immunostaining in distinguishing small cell lung carcinomas from other small cell carcinomas. *Am J Surg Pathol* 2000;24:1217-23.
- 80. Ralston J, Chiriboga L, Nonaka D. MASH1: a useful marker in differentiating pulmonary small cell carcinoma from Merkel cell carcinoma. *Mod. Pathol* 2008;21:1357–62.
- 81. Hiroshima K, Iyoda A, Shida T, et al. Distinction of pulmonary large cell neuroendocrine carcinoma from small cell lung carcinoma: a morphological, immunohistochemical, and molecular analysis. *Mod Pathol* 2006;19:1358–68.
- 82. Shuda M, Arora R, Kwun HJ, et al. Human Merkel cell polyomavirus infection I. MCV T antigen expression in Merkel cell carcinoma, lymphoid tissues and lymphoid tumors. *Int J Cancer* 2009;125:1243-9.
- 83. Koljonen V, Tukiainen E, Haglund C, et al. Proliferative activity detected by Ki67 correlates with poor outcome in Merkel cell carcinoma. *Histopathology* 2006;49:551–3.
- 84. Carson HJ, Reddy V, Taxy JB. Proliferation markers and prognosis in Merkel cell carcinoma. *J Cutan Pathol* 1998;25:16-9.
- 85. Fernández-Figueras MT, Puig L, Musulén E, et al. Expression profiles associated with aggressive behavior in Merkel cell carcinoma. *Mod Pathol* 2007;20:90-101.
- 86. Asioli S, Righi A, Volante M, Eusebi V, Bussolati G. p63 expression as a new prognostic marker in Merkel cell carcinoma. *Cancer* 2007;110:640-7.
- 87. Altieri D. Survivin, versatile modulation of cell division and apoptosis in cancer. Oncogene 2003;22:8581.
- 88. Kim J, McNiff JM. Nuclear expression of survivin portends a poor prognosis in Merkel cell carcinoma. *Mod Pathol* 2008;21:764–9.
- 89. Andea AA, Patel R, Ponnazhagan S, et al. Merkel cell carcinoma: correlation of KIT expression with survival and evaluation of KIT gene mutational status. *Hum Pathol* 2010;41:1405-12.
- 90. Fernandez-Figueras MT, Puig L, Musulen E et al. Expression profiles associated with aggressive behavior in Merkel cell carcinoma. *Mod Pathol* 2007;20:90–101.

- 91. Penneys NS, Shapiro S. CD44 expression in Merkel cell carcinoma may correlate with risk of metastasis. J Cutan Pathol 1994;21:22-6.
- 92. Woegerbauer M, Thurnher D, Houben R, et al. Expression of the tetraspanins CD9, CD37, CD63, and CD151 in Merkel cell carcinoma: strong evidence for a posttranscriptional fine-tuning of CD9 gene expression. Mod Pathol 2010;23:751-62

Conflict de interese NEDECLARATE

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